**Supplementary Materials**

**DNA oligonucleotide primer list:**

AD49 5’ TTCACTGACAGCTGTGAGCTACAC 3’

AD50 5’ CAGCGGAGCATTACAGAACGACAG 3’

AD57 5’ GTGTTAGACTGAACATTTTAGAG 3’

AD58 5’ TTTGCCATTCTCCGAAGTTGATG 3’

Cbtra-2RTCF – 5’ CGAGGTTTTATCACTGGTC 3’

Cbtra-2RTCR-- 5’ GATGCTCTCCCAGGATGAT 3’

Cbtra-2GENINCFO -- 5’GATCGGGTAGTCACCTATCTAAC 3’

Cbtra-2GENINRE -- 5’ CAGCAATGAGGAATGCAGGTAGA 3’

DK1 – 5’ GACTGATCTTCTGGAGGTTAACGG 3’

DK2 – 5’ TTCGGAATATCGGAATCTCAGGA 3’

DK3 – 5’ TGCTTGCGGGCAGAACTACTTCCA 3’

DK4 – 5’ GTCATTCCGACTTCGTCAACGGTT 3’

DK5 – 5’ GGCCTTTGAATTCTCAGGTTCTTG 3’

DK6 – 5’ GGAGATCAAGACTCTGAAGTTTGG 3’

DK7 – 5’ ACACTTTCAGATGATGATGGCTGC 3’

DK8 – 5’ CAGGCCTTTTCTCTAGGTCAAGTT 3’

DK9 – 5’ CTAGGCCATTTTGACATGGTAGCC 3’

DK10 – 5’ TCCTCCCTATTAGGCCTTGGTGGT 3’

DK11 – 5’ TCGCAGTCTTCAATTCCCTCGCCG 3’

DK12 – 5’ TACGAAGCACTATGCTCCACGGCC 3’

DK13 – 5’ GATTCCATCACAGACCTGATTCGG 3’

DK14 – 5’ CCGAATCAGGTCTGTGATGGAATC 3’

DK15 – 5’ CGAGAAGGAGACGCTGAACTGTCC 3’

DK16 – 5’ GGACAGTTCAGCGTCTCCTTCTCG 3’

DK17 – 5’ CGCCTCTGGACATGATCCTTGTGT 3’

DK18 – 5’ ACACAAGGATCATGTCCAGAGGCG 3’

DK19 – 5’ GTCGTGTCAGAGGAGCTGTCAGGT 3’

DK20 – 5’ ACCTGACAGCTCCTCTGACACGAC 3’

DK21 – 5’ CATCATCCTGGGAGAGCATCAACG 3’

DK22 – 5’ CGTTGATGCTCTCCCAGGATGATG 3’

DK23 – 5’ GAAGCTCACCCGTGCCGTCAACTT 3’

DK24 – 5’ AAGTTGACGGCACGGGTGAGCTTC 3’

DK25 – 5’ GAAGGGCCCCCTTCCAAATATGTA 3’

DK26 – 5’ CGACAGCGGTGTGCCGATTCCACC 3’

DK27 – 5’ CGTCTTCTGACTGGTCTACGTGAG 3’

DK28 – 5’ TGGTTCCGATGTGACGCGGAGCCG 3’

DK29 – 5’ GAGCTCTCGGGCATCTGAAAGCAC 3’

DK30 – 5’GCCAGTCGGCTGGCGCCTTTGTTT 3’

DK31 – 5’ TGTGTGTGCGTTGGCTCCGCCCAC 3’

DK32 – 5’ GCGGCCGTCGGATTGCATACATAA 3’

DK33 – 5’GGAGGAGGTGTGAGTGTGAGAGGG 3’

DK34 – 5’ TTGCCGCACCAGGTGTTCATCAGC 3’

DK35 – 5’ GCTGCAGGAACGGATGTCGGGATG 3’

DK36 – 5’ CGCCGCTTACACCAATGGGAGCAA 3’

DK37 – 5’ TCATCGTCGAAGGCTTCAGCAGCC 3’

DK38 – 5’ GGTGGACGCCCGATCCTGTCGGTT 3’

DK39 – 5’ GCGGCACGAACAATGCGATGATGA 3’

DK40 – 5’ TGCTGAAGCAGTTCCCGGAACATC 3’

DK41 – 5’ GAAGGAGGGGGCGAGAGCCGTTGA 3’

DK42 – 5’ GAGCATCGCCGAACGATGTTACTC 3’

DK45 – 5’ CTGTCGGGTCCTGAAAGCATACAG 3’

DK48 – 5’ CACTGACAGCTGTGAGCTACACGG 3’

DK49 – 5’ CAGAGCTGTCCCGACCATTTGTCG 3’

EB2 – 5’ CGAAGCTACCTCCGGATAGCCC 3’

EB3 -- 5’ CGTTCGGCTCTTTATCTAAC 3’

EH23 -- 5’ TAATACGACTCACTATAGGGCGCTGCTCTGATGGATCCGAATG 3’

EH24 – 5’ TAATACGACTCACTATAGGGCCGTTGATTACGGTGCGAGCCAC 3’

EH29 -- 5’ GCCTGGGCTGGCTAATGGTTC 3’

EH30 -- 5’ CTTGCTGCAGTTGGTTCCGAAG 3’

EH33 – 5’ GGATTGAGCCATTAAATGGTCG 3’

EH34 – 5’ TTTAGCTTCATCCAATTAAATC 3’

RE883 – 5’ GGACGTACCACCGGAGTCGTCC 3’

RE884 – 5’ GCATACGATCAGCAATTCCTGGG 3’

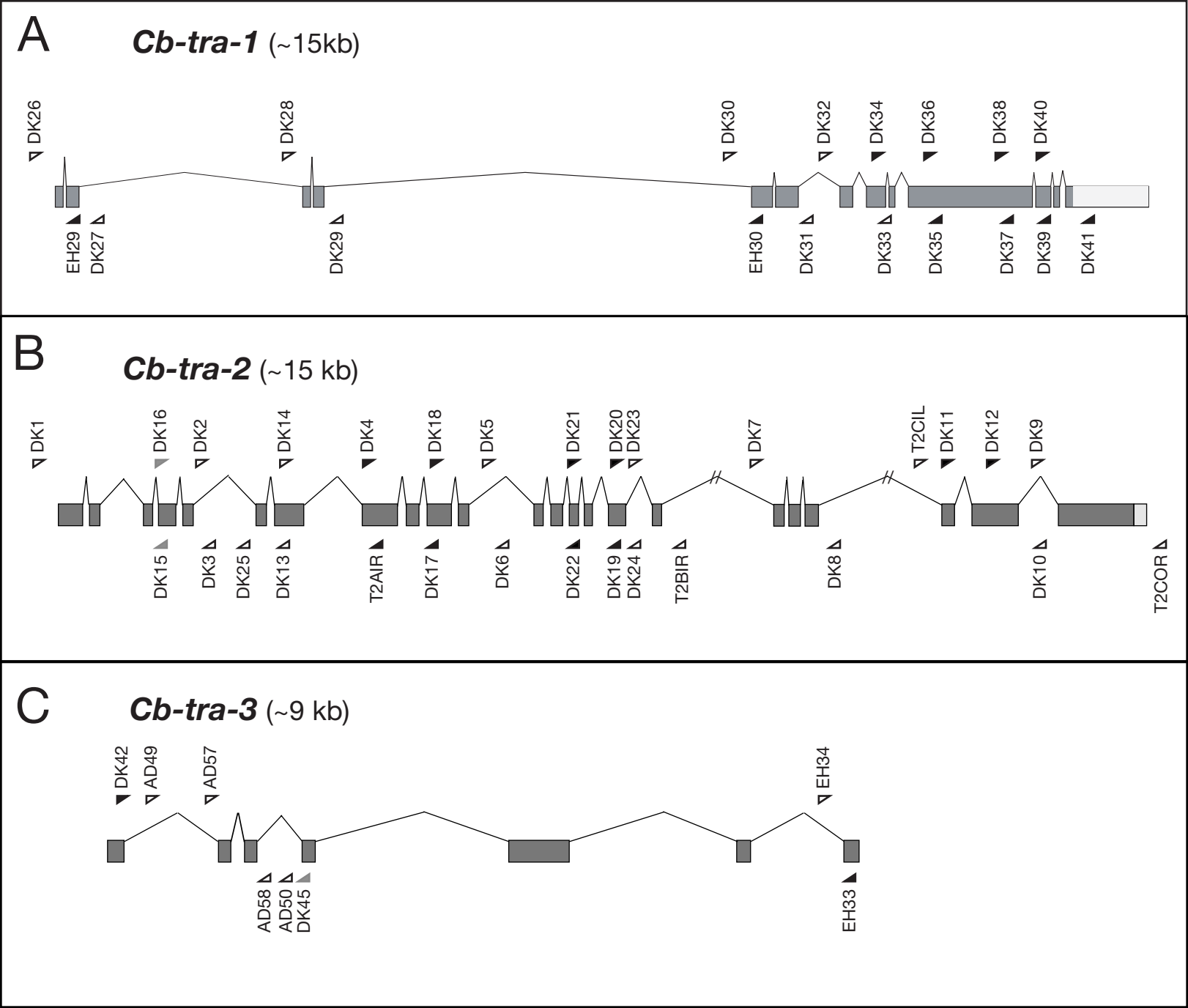
T2AIR — 5’ CTTCTACTGGATCGACTCTCGC 3’

T2BIR — 5’ GATAAAGGAAGTGTGGGTCGAG 3’

T2CIL — 5’ CCTCGTGGACAATTCGATGTGCG 3’

T2COR— 5’ CTGTAACTGCCTCCATTATGCG 3’

**Figure S1. Locations of primer annealing sites.** Intron-exon diagrams for *Cb-tra-1* (A, DE BONO AND HODGKIN 1996), *Cb-tra-2* (B, KUWABARA, 1996), and *Cb-tra-3* (C, WormBase prediction CBG21580, generally verified by this work) are shown, with the approximate locations of various primers indicated. Wedges point in the 5' to 3' direction, such that those above the splicing diagram are sense relative to transcription, and those below are antisense. Wedges filled with black lie entirely in exonic sequences and those that are open are entirely intronic. Primers that straddle an intron-exon junction are shown in gray.


A
Cb-tra-1 (~15kb)
DK26
DK28
DK30
DK32
DK34
DK36
DK38
DK40
EH29
DK27
DK29
EH30
DK31
DK33
DK35
DK37
DK39
DK41
B
Cb-tra-2 (~15 kb)
DK1
DK16
DK2
DK14
DK4
DK18
DK5
DK21
DK20
DK23
DK7
T2CIL
DK11
DK12
DK9
DK15
DK3
DK25
DK13
T2AIR
DK17
DK6
DK22
DK19
DK24
T2BIR
DK8
DK10
T2COR
C
Cb-tra-3 (~9 kb)
DK42
AD49
AD57
EH34
AD58
AD50
DK45
EH33